

## AMENDMENTS TO THE SPECIFICATION

The following stated paragraphs replace all prior versions of these paragraphs in the application.

[0050] The present invention can be practiced in any of the usual enzymatic reaction formats. Thus, a dye labeled peptide substrate is first prepared, being selected according to desired specificity for the enzyme of choice. The labeled peptide may utilize any of the recognized fluorophores as the dye. Examples of dyes considered to be useful include Lissamine Rhodamine, BODIPY dyes (Molecular Probes, Inc., Eugene, Oreg.), fluorescein, and Oregon Green. Other examples of fluorescent dyes are other dyes supplied by Molecular Probes, as well as those fluorescent dyes manufactured by Amersham and Dyomics and others. Preparation of the dye labeled peptide substrate is accomplished by commonly known procedures. For example, attachment of the fluorophore to the peptide sequence is conveniently accomplished during peptide synthesis by reaction of the N-terminus amino group of the peptide with the dye. A nucleophilic electrophilic, reactive derivative of the fluorophore, such as a sulfonyl chloride derivative, may be utilized to effect the covalent attachment of the fluorophore to the N-terminal alpha amino group of the peptide. However, other methods of attachment and at other locations can be utilized.

[0080] Okadaic Acid, another phosphatase inhibitor, solubilized in DMSO, was titrated in triplicate in phosphatase dilution buffer over 24 wells in white 96-well polystyrene COSTAR ~~G~~star plate. PP2A Phosphatase enzyme and Lissamine Rhodamine labeled phospho Kemptide peptide substrate in Example VII were added to a final concentration of 22 milli Units and 30  $\mu$ M, respectively. The final incubation mixture in the well was 30  $\mu$ l. The reaction was stopped at 60 minutes with the addition of the Working Solution prepared according to Example I and then diluted with 30 volumes of water. Fluorescence was measured using a BMG FluoStar plate reader with a 560/590 ex/em filter set. The results are illustrated in FIG. 10. The calculated IC<sub>50</sub> was found to be 1.5 nM.